# Diagnostic value of calprotectin in differentiation between benign and malignant pleural effusion

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Background Pleural effusion can arise as a result of more than 50 recognized causes and the differentiation between benign and malignant origin of the fluid is still a diagnostic challenge. The ability of tumor markers and other biological markers to make better diagnosis of malignant pleural effusion (MPE) remains questionable. Out of these, the calcium-related proteins S100-A8 and S100-A9 (the noncovalent heterodimer calprotectin) were demonstrated in a small amount in malignant not in benign pleural effusion.

Objectives This research aimed to assess the diagnostic value of calprotectin in the differentiation between infectious or benign and MPE.

Patients and methods Sixty patients were divided into group I: malignant and group II: infectious pleural effusions (which were further divided into group IIA: parapneumonic effusion and group IIB: tuberculous effusion) Quantitative measurement of calprotectin was done using the enzymelinked immunosorbent assay technique in pleural effusion.

Results Pleural calprotectin level in MPEs (229.2±168.6 ng/ ml) was significantly lower than its level of infectious pleural

effusions (3202.2±1304.8 ng/ml; P<0.001). The cutoff value of calprotectin level for the diagnosis of MPE was less than or equal to 730.5 ng/ml, with 95% confidence interval and the area under the curve was 0.999, the corresponding sensitivity was 96.7 and the specificity was 100% (P<0.001).

Conclusion Calprotectin is a valuable biomarker in differentiating malignant from infectious pleural effusion. Egypt J Bronchol 2019 13:382-387 © 2019 Egyptian Journal of Bronchology

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Keywords: calprotectin, malignant pleural effusion, nonmalignant pleural effusion, pleural effusion

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# Introduction

Diagnosis of pleural effusion remains a challenge. The list of the diseases they cause is big as well as it is heterogeneous. The more important difficulty in the diagnosis of exudative effusions is to differentiate benign from malignant effusion [1]. Most of malignant pleural effusions (MPEs) (90-97%) are exudative; they result from increased filtration from pleural vessels [2]. The initial semi-invasive method to diagnose exudative pleural effusion (thoracocentesis) allows cytological, microbiological, and biochemical analyses of the fluid [3]. More invasive procedures are required to diagnose exudative effusion with negative cytology particularly if malignancy is suspected. Closed pleural biopsy has a little diagnostic value and because of high diagnostic yield of thoracoscopy ( $\geq$ 90%), it is the method of choice [4].

Many studies have assessed the capability of tumor markers and other biological markers to make a better diagnosis of MPE [5]. Several proteins were demonstrated in malignant versus benign pleural effusion (BPE). Out of these, the calcium-related proteins S100-A8 and S100-A9 (noncovalent heterodimer calprotectin) were demonstrated in a small amount in MPE [6].

Calprotectin is a calcium- and zinc-binding protein of the S100 group heterodimeric complex [7]. Magne

Fagerhol and colleagues first described calprotectin in 1980 [8], it is a 36 kDa protein with two 14 kDa and one 8 kDa chain of amino acid, these proteins were named so because they were 100% soluble in ammonium sulfate solution [9]. Calprotectin is heat resistant and resistant to proteolysis in the presence of calcium [10]. When neutrophil activation or endothelial adhesion of monocytes calprotectin begins to be secreted by a microtubulemediated alternative pathway, thus acting as a marker for the influx of phagocytes into the site of inflammation [11],leading increase concentration in the plasma, serum, spinal fluid, synovial fluid, pleural fluid, urine, saliva, and stool during bacterial infection or inflammation in the relevant organs [12]. It has bacteriostatic and fungistatic properties that arise from its ability to sequester manganese and zinc [13]. It causes inhibition of metalloproteinases and chelation with zinc and manganese ions to inhibit microbial proliferation, as these metals are of vital importance for bacterial growth [14]. Calprotectin induces the

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apoptosis both in malignant and nonmalignant cell lines [15].

The function of calprotectin in cancer biology is conflicting to some extent. It is a strong apoptotic factor when produced by immune cells. On the contrary, expression in cancer cells is related to tumor development, cancer invasion, and metastasis [9]. Its relationship with inflammation is obvious with an established proinflammatory role in various inflammatory states [16]. A high calprotectin level could be expected in a patient with inflammatory disorder [15]. It could be reasonably expected to find high quantities of this protein in MPE because it is also concerned with inflammation-associated carcinogenesis [9].

The higher levels of calprotectin found in BPE could attribute to the antimicrobial role of this protein. When neutrophil dies as a policy to suppress the growth of various fungal and bacterial pathogens, a massive amount of calprotectin is released [17]. Accordingly, calprotectin levels were high in parapneumonic and tuberculous pleural effusion and low in additional benign noninfectious etiologies [18].

## Aim of the work

The objective of this study is to clarify the diagnostic value of calprotectin in the differentiation between infectious or benign and MPE.

#### Patients and methods

This study was conducted on 60 patients with pleural effusion admitted at the Chest Department, Benha University Hospital during the period from December 2016 till January 2018. Ethical research approval from Benha University Hospitals ethics committee and informed consent from the patient were obtained.

The patients were classified according to their final diagnosis into two groups.

- (1) Group I: 30 patients with MPE.
- (2) Group II: patients with infectious pleural effusion who were subdivided into two groups:
  - (a) Group IIA, 15 patients with parapneumonic pleural effusion.
  - (b) Group IIB, 15 patients with tuberculous pleural effusion.

Patient with any of the following criteria was excluded [19]:

(1) Under treatment with anticancer chemotherapy.

- (2) Under treatment with antituberculous drugs.
- (3) Using glucocorticoid and other anti-inflammatory drugs.

All patients were subjected to the following:

- (1) Thorough medical history: age, sex, residence, occupation, smoking, and other particular habits of medical importance.
- (2) General examination.
- (3) Local chest examination.
- (4) Routine laboratory investigations in the form of:
  - (a) Complete blood picture.
  - (b) Erythrocyte sedimentation rate.
  - (c) Coagulation profile.
  - (d) Fasting and 2 h postprandial blood glucose.
  - (e) Liver function tests.
  - (f) Kidney function tests: to exclude renal failure.
- (5) Radiological examination including plain chest radiography posteroanterior and lateral views and computed tomography (CT) scans of the chest.
- (6) Sputum examination for acid-fast alcohol fast bacilli by Ziehl-Neelsen stain.
- (7) Tuberculin skin test [20]: using the Mantoux method where 0.1 ml (5 tuberculin units) of purified protein derivative was intradermally into the volar aspect of the forearm, then the test was read 48-72 h later, the reaction was considered positive if indurations of 10 mm or more in diameter were detected.
- (8) Diagnostic thoracocentesis: the obtained pleural fluid (about 300-500 ml) was subjected to the following examinations:
  - (a) Physical examination, including color, aspect, turbidity, and specific gravity.
  - (b) Chemical examination, including proteins, glucose, and lactate dehydrogenase (LDH), where effusions were classified into exudates or transudates according to Light's criteria. If the effusion had any of the following three properties, the effusion was classified into exudates:
    - (i) A ratio of the concentration of total proteins in pleural fluid to serum total proteins of more than 0.5.
    - (ii) An absolute value of LDH of more than 200 IU.
    - (iii) A ratio of pleural fluid LDH to serum LDH of more than 0.6 [20].
  - (c) Bacteriological examination: including culture and sensitivity, pleural examination for Gram stain, to exclude infectious causes other than tuberculosis,

- and for acid-fast alcohol fast bacilli to detect tuberculosis bacilli by direct smear examination.
- (d) Cytological examination: using the Papanicolaou-stained smears, hematoxylin and eosin-stained section of paraffinembedded cell blocks.
- (9) Pleural biopsies: were taken for all patients in groups (I, II) by either of the following methods: closed pleural biopsies using Abram's needle or thoracoscopic biopsies.
- (10) Assay of the soluble calprotectin level in the pleural fluid by enzyme-linked immunosorbent assay: the principle of the test; the kit used double-antibody sandwich enzyme-linked immunosorbent one-step process assay to assay the level of calprotectin in pleural effusion. Add standard, the test sample and horseradish peroxidase-labeled calprotectin in pleural effusion antibodies to enzyme wells which are precoated with calprotectin in pleural effusion antibody, then incubation was carried out and washed to remove the uncombined enzyme. Upon chromogen solutions A and B, the color of the liquid changed to blue, and the reaction with the acid caused the color to become yellow. The depth of color and the concentration of the calprotectin in pleural effusion sample were positively correlated.

## Statistical analysis [21]

The collected data were tabulated and analyzed using SPSS, version 16 software (SPSS Inc. Released 2007, SPSS for Windows, version 16.0; SPSS Inc., Chicago, Illinois, USA). Categorical data were presented as number and percentages, using  $\chi^2$  test to analyze them. Quantitative data were expressed as a mean ±SD, median, interquartile range. They tested for normality using the Shapiro-Wilk test, assuming normality at a P value of more than 0.05, Student's t test, and analysis of variance for normally distributed variables. Nonparametric variables were analyzed using Mann-Whitney, Wilcoxon test, the Kruskal-Wallis test. Receiver operating characteristic curve was used to detect the cutoff value of calprotectin in the prediction of MPE.

The degree of significance in this work started below 0.05 (P<0.05 was considered significant). A P value of more than 0.05 is nonsignificant (NS), P value less than 0.05 is significant (S), and a P value less than or equal to 0.001 is highly significant (HS).

#### Results

This study included 60 patients: 26 (43.3%) men and 34 (56.4%) women. There were 14 (46.7%) men and 16 (53.3%) women in group I (malignant effusions) and there were 16 (53%) men and 14 (47%) women in group II (infectious effusions) which were subdivided into group IIA: the parapneumonic effusion group had 10 (67 %) men and five (33 %) women while group IIB included six (40%) men and nine (60%) women with tuberculous pleural effusion without significant difference between them.

The ages of the studied patients ranged from 19 to 76 years: group I (malignant effusions) with a mean age of 57.6±13 while group II (infectious effusions) was subdivided into group IIA, parapneumonic pleural effusion (their ages ranged 29-75 years with a mean 52±14) and group IIB, tuberculous pleural effusion (their ages ranged 13-65 years with a mean 44.8 ±14.5) with a significant difference tuberculous and MPE.

The pleural calprotectin in group I MPE (229.2 ±168.6 ng/ml) was significantly lower than its level in group II; infectious pleural effusion was (3202.2) ±1304.8 ng/ml) with highly significant difference between two groups (Table 1).

The patients suffered from parapneumonic pleural effusions, group IIA demonstrated the highest levels of calprotectin (3333 ng/ml) and its level in patients with tuberculous pleural effusions group IIB (3071.3 ng/ml) were significantly higher than those of MPE group I (229.2 ng/ml), P value less than 0.001 (HS) (Table 2). However, the pleural calprotectin level in parapneumonic pleural effusion group IIA (3333±1151.8 ng/ml) was not substantially different from those in the tuberculous pleural effusion group IIB (3071.3 ng/ml) (P=0.92) (Table 3 and Fig. 1).

Table 1 Statistical analysis of pleural calprotectin level among the two studied groups

| Effusion     | N  | Ple           | ural calprotectin |        | MWU  | Р           |  |
|--------------|----|---------------|-------------------|--------|------|-------------|--|
|              |    | Mean±SD       | Min               | Max    |      |             |  |
| Malignant    | 30 | 229.2±168.6   | 99.0              | 1040.0 | 6.65 | <0.001 (HS) |  |
| Nonmalignant | 30 | 3202.2±1304.8 | 1040.0            | 5400.0 |      |             |  |

HS, highly significant; Max, maximum; Min, minimum; MWU, Mann-Whitney U test.

Table 2 Statistical analysis of pleural calprotectin level among the studied subgroup

| Groups        | Ν  | Pleural calprotectin |         |         | KWT  | Р           | Significant pairs                |
|---------------|----|----------------------|---------|---------|------|-------------|----------------------------------|
|               |    | Mean±SD              | Minimum | Maximum |      |             |                                  |
| Malignant     | 30 | 229.2±168.6          | 99.0    | 1040.0  | 44.1 | <0.001 (HS) | TB≠Malignant Parapneu.≠malignant |
| Parapneumonic | 15 | 3333.0±1151.8        | 1040.0  | 5333.0  |      |             |                                  |
| Tuberculous   | 15 | 3071.3±1470.8        | 1145.0  | 5400.0  |      |             |                                  |

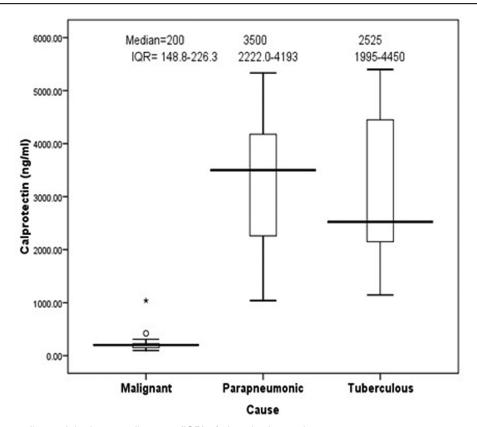
HS, highly significant; KWT, Kruskal-Wallis test; TB, tuberculosis.

Table 3 Statistical analysis of pleural calprotectin level in tuberculous and parapneumonic groups

| Effusion      | N  | Ple           | ural calprotectin | MWU    | P     |           |
|---------------|----|---------------|-------------------|--------|-------|-----------|
|               |    | Mean±SD       | Min               | Max    |       |           |
| Tuberculous   | 15 | 3071.3±1470.8 | 1145.0            | 5400.0 | 0.104 | 0.92 (NS) |
| Parapneumonic | 15 | 3333.0±1151.8 | 1040.0            | 5333.0 |       |           |

Max, maximum; Min, minimum; MWU, Mann-Whitney U test.

Figure 1



Box plot showing the median and the interquartile range (IQR) of pleural calprotectin.

Table 4 Cutoff value, sensitivity, specificity, and positive predictive value of pleural calprotectin level in the prediction of malignant pleural effusion

| Variables            | Cutoff | Sens% | Spec% | PPV% | NPV% | Accuracy% | AUC   | 95% CI   | Р           |
|----------------------|--------|-------|-------|------|------|-----------|-------|----------|-------------|
| Pleural calprotectin | ≤730.5 | 96.7  | 100   | 100  | 96.8 | 98.3      | 0.999 | 0.99-1.0 | <0.001 (HS) |

AUC, area under the curve; CI, confidence interval; HS, highly significant; NPV, negative predictive value; PPV, positive predictive value; Sens, sensitivity; Spec, specificity.

The cutoff value of calprotectin level for the diagnosis of MPE was less than or equal to 730.5 ng/ml; therefore, patients with calprotectin concentrations lower than this threshold had a high possibility of being diagnosed as MPE. The value for the area

under the corresponding receiver operating characteristic curve [area under the curve (AUC)] was 0.999, the 95% confidence interval was 0.91-1 with 96.7% sensitivity and 100% specificity (*P*<0.001) (Table 4).

#### **Discussion**

Diagnosis of pleural effusion remains a challenge and the more important difficulty in the diagnosis of exudative effusion is to differentiate benign from malignant effusion [1].

This current study was carried out on 60 patients with exudative pleural effusion [group I with the mean age of 57.6±13 and group II (tuberculous with mean age 44.8±14.5, parapneumonic with mean age 52±14)].

These results are in accordance with Luo et al. [22] who studied 95 patients with pleural effusion; their mean age was as follows; malignant group 61±13.1; infectious group 47±20.6 (tuberculous 45±20.6). Sanchez-Otero et al. [18] also carried out a study on 156 patients, with 67 patients in the MPE group (men and 25 women, 42.9%) 42 and 89 patients in the BPE group (57.05%), where patients with BPE were secondary to tuberculosis (n=30, 20 men, 10 women) and parapneumonic (n=29, 22 men, seven women) and their mean age was 67.00±13.9 in MPE and 53±20 in BPE.

The pleural calprotectin in MPE (229.2±168.6 ng/ml) was significantly lower than its level in infectious pleural effusion (3202.2±1304.8 ng/ml) (Table 1). Sanchez-Otero et al. [18] had studied concentration of pleural fluid calprotectin in the different diagnostic categories. The mean level of calprotectin in patients with MPE (257.2 ng /ml) was significantly lower than those with BPE (2627.1 ng/ml). Kohmo *et al.* [23] compared calprotectin and CXCL12 in MPE with that in BPE and demonstrated that calprotectin and the CXCL12 level significantly increased nonmalignant pleural fluid compared with malignant pleural fluid, which supported the measurement of both biomarkers in pleural effusion as a possible noninvasive strategy for the differential diagnosis of MPE. Also Lou et al. [22] found that the median level of calprotectin and CXCL12 in MPE were 447.15 and 4.12 ng/ml, and both were significantly lower than that of the BPE (P=0.003 and 0.020, respectively) and that of tuberculous pleural effusion (P=0.002 and 0.003, respectively). The higher calprotectin levels found in BPE could attribute to the antimicrobial role of this protein [17].

Calprotectin level in patients with parapneumonic and tuberculous pleural effusions were significantly higher than those with MPE [3333, 3071.3, and 229.2 ng/ml, respectively with P<0.001 (HS)] (Table 2). However, the pleural calprotectin level in parapneumonic pleural effusion was not significantly different from those in

the tuberculous pleural effusion (P=0.92) (Table 3). Sanchez-Otero et al. [18] found that patients with pneumonia showed the highest calprotectin level (3517.9 ng/ml) and were significantly higher than those of the MPF group (257.2 ng/ml), but these levels were not considerably different from those in the tuberculous group (2982.3±1573.0 ng/ml).

In the present work, the cutoff value of calprotectin level for the diagnosis of MPE was less than or equal to 730.5 ng/ml with 95% confidence interval and the AUC was 0.999; the corresponding sensitivity was specificity was 100% (P < 0.001)96.7% and (Table 4). Sanchez-Otero and colleagues detected the ability of calprotectin to differentiate between MPE and BPE by initiating two cutoff points. The first one (≤545 ng/ml) was determined as it gave the highest accuracy level (92.31%), and a sensitivity of 97.01% and specificity of 88.76%, while the second one (≤736.4 ng/ml) represented a sensitivity of 100%, specificity of 83.1%, accuracy of 90.4%, and AUC was found to be 0.963 (95% confidence interval, 0.932–0.994). Besides, they analyzed the calprotectin capability to differentiate pleural effusion subtypes. It showed the highest accuracy in differentiating MPE pleural effusion of tuberculous from and (97.94)parapneumonic origin and 95.83%, respectively) [18]. Luo et al. [22] stated that high calprotectin showed specificity discriminating MPE from BPE and tuberculous pleural effusion (71.43 and 84.09%, respectively). On the contrary, Su and colleagues found a strong expression of S100-A8, S100-A9 adenocarcinoma and end-stage lung cancer tissue. Also, Blanco-Prieto and colleagues found a similar level of calprotectin in the sera of lung cancer patients (221.21 ng/ml) but a lower serum level was found in benign inflammatory conditions (141.93 ng/ ml). They explained that finding by the different nature of the fluids (pleural vs. serum) and their higher expression in lung tissues may be explained by the combined effect of inflammation progression [24,25].

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### **Conflicts of interest**

There are no conflicts of interest.

## References

1 Mohamed SA, Agmy GR, Wafy SA, Abd El-Hameed MG. Value of Creactive protein in differentiation between tuberculous and malignant pleural effusion. Egypt J Bronchol 2017; 11:49-55.

- 2 Neragi-Miandoab S. Malignant pleural effusion, current and evolving approaches for its diagnosis and management. Lung Cancer 2006; 54:1-9.
- 3 Hooper C, Lee YC, Maskell N. Investigation of unilateral pleural effusion in adults: British Thoracic Society Pleural Disease Guideline. Thorax 2010; 65 (Suppl 2):ii4-ii17.
- 4 Roberts ME, Neville E, Berrisford RG, Antunes G, Ali NJ. Management of malignant pleural effusion: British Thoracic Society Pleural Disease Guideline 2010. Thorax 2010; 65:ii32-ii40.
- 5 Botana-Rial M, Casado-Rey P, Leiro-Fernandez V, Andrade-Olivie M, Represas-Represas C, Fernandez-Villar A. Validity of procalcitonin and C-reactive protein measurement when differentiating between benign and malignant pleural effusion. Clin Lab 2011; 57:373-378.
- 6 Rodriguez-Pineiro AM, Blanco-Prieto S, Sanchez-Otero N, Rodriguez-Berrocal FJ, de la Cadena MP. On the identification of biomarkers for nonsmall cell lung cancer in serum and pleural effusion. J Proteomics 2010; 73:1511-1522.
- 7 Vermeire S, Van Assche G, Rutgeerts P. Laboratory markers in IBD: useful, magic, or unnecessary toys? Gut 2006; 55:426-431.
- 8 Fagerhol MK, Dale I, Andersson T. Release and quantitation of a leukocyte derived protein L1. Scan J Haematol 1980; 24:393-398.
- 9 Ghavami S, Chitayat S, Hashemi M, Eshraghi M, Chazin WJ, Halayko AJ, et al. S100A8/A9: a Janus-faced molecule in cancer therapy and tumorigenesis. Eur J Pharmacol 2009; 625:73-83.
- 10 Odegaard E, Davidson B, Engh V, Onsrud M, Staff AC. Assessment of endoglin and calprotectin as potential biomarkers in ovarian carcinoma and borderline tumors of the ovary. Obstet Gynecol 2008; 199:533.el-8.
- 11 Hermann A, Hess J, De Servi B, Medunjanin S, Grobholz R, Trojan L, et al. Calcium-binding proteins S100A8 and S100A9 as novel diagnostic markers in human prostate cancer. Clin Cancer Res 2005; 11:5146-5152.
- 12 Kremer R. Best LA. Savulescu D. Gavish M. Nagler RM. Pleural fluid analysis of lung cancer vs. benign inflammatory disease patients. Br J Cancer 2010; 102:1180-1184.
- 13 Zhang W, Yang HC, Wang Q, Yang ZJ, Chen H, Wang SM, et al. Clinical value of combined detection of serum matrix metalloproteinase-9, heparanase, and cathepsin for determining ovarian cancer invasion and metastasis. Anticancer Res 2011: 31:3423-3428.

- 14 Srikrishna G, Panneerselvam K, Westphal V. Two proteins modulating transendothelial migration of leukocytes recognize novel carboxylated glycans on endothelial cells. J Immunol 2001; 166:4678-4688.
- 15 Kallel L, Fekih M, Boubaker J, Filali A. Faecal calprotectin in inflammatory bowel diseases: a review. Tunis Med 2011: 89:425-429.
- 16 Gebhardt C, Nemeth J, Angel P, Hess J. S100A8 and S100A9 inflammation and cancer. Biochem Pharmacol 2006;
- 17 17.Nisapakultorn K, Ross KF, Herzberg MC. Calprotectin expression inhibits bacterial binding to mucosal epithelial cells. Infect Immun 2001; 69:3692-3696.
- 18 Sanchez-Otero N, Blanco-Prieto S, Paez de la Cadena M, Vázquez-Iglesias L, Fernández-Villar A, Botana-Rial MI, et al. Calprotectin: a novel biomarker for the diagnosis of pleural effusion. Br J Cancer 2012;
- 19 Xuan W, Zhang J, Zhou Q, Ma LJ. Role of interleukin -33 in the differentiation between tuberculous and malignant pleural effusion. Oncol Lett 2014; 8:449-453.
- 20 Light RW. Pleural diseases. 4th ed. Philadelphia, PA: Lippincott Williams, and Wilkins; 2001. 392-394
- 21 Kothari CR. Research methodology: methods and techniques. 2nd ed. New Delhi: New Age International Publishers; 2004.
- 22 Luo J, Wang M, Li C, Liang B, Liu D, Shi C, et al. A novel combination of calprotectin and CXCL12 for predicting malignancy in patients with exudative pleural effusion. Medicine (Baltimore) 2015; 94:e2105.
- 23 Kohmo S, Kijima T, Mori M, Minami T, Namba Y, Yano Y, et al. CXCL12 as a biological marker for the diagnosis of tuberculous pleurisy. Tuberculosis 2012: 92:248-252.
- 24 Su YJ, Xu F, Yu JP, Yue DS, Ren XB, Wang CL. Up-regulation of the expression of S100A8 and S100A9 in lung adenocarcinoma and its correlation with inflammation and other clinical features. Chin Med J 2010; 123:2215-2220.
- 25 Blanco-Prieto S, Vázquez-Iglesias L, Rodriguez-Girondo M, Barcia-Castro L, Femández-Villar A, Botana-Rial MI, et al. Serum calprotectin, CD26 and EGF to establish a panel for the diagnosis of lung cancer. PloS ONE 2015; 10:e0127318.